

# Molecular placement of experimental electron density: a case study on UDP-galactopyranose mutase

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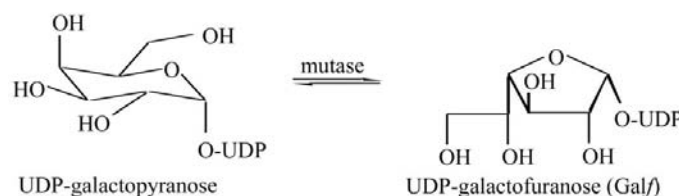
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The structure of UDP-galactopyranose mutase, the enzyme responsible for the conversion of UDP-galactopyranose to UDP-galactofuranose, has been solved. The structure solution required the use of two crystal forms and a selenomethionine variant. Crystal form  $P2_1$  was used to collect a complete MAD data set, a native data set and a single-wavelength non-isomorphous selenomethionine data set. A starting set of MAD phases was then improved by non-crystallographic averaging and cross-crystal averaging of all  $P2_1$  data. The initial maps were of such low quality that transformation matrices between cells could not be determined. It was therefore assumed that although there were large changes in unit-cell parameters, the molecule occupied the same position in each cell. This starting assumption was allowed to refine during the averaging procedure and did so satisfactorily. Despite a visible increase in the quality of the map allowing some secondary-structural elements to be located, the overall structure could not be traced and refined. The rediscovery of the second crystal form,  $P2_12_12_1$ , allowed the collection of a native data set to 2.4 Å. Molecular placement of electron density was used to determine the relationship between the two unit cells. In this study, only the already averaged  $P2_1$  experimental density could be placed in the  $P2_12_12_1$  map. Less extensively density-modified maps did not give a clear solution. The study suggests even poor non-isomorphous data can be used to significantly improve map quality. The relationship between  $P2_1$  and  $P2_12_12_1$  could then be used in a final round of cross-crystal averaging to generate phases. The resulting map was easily traced and the structure has been refined. The structure sheds important light on a novel mechanism and is also a therapeutic target in the treatment of tuberculosis.

## 1. Introduction

We have previously reported the purification and crystallization of mutase from *Escherichia coli* (McMahon *et al.*, 1999). Mutase initially crystallized in two different forms, only one of which was reproducible. These crystals (in the  $P2_1$  space group) were small two-dimensional diamond-shaped crystals that proved very difficult to work with, as they diffracted poorly and were highly mosaic. This paper describes the rediscovery of the second crystal form of mutase (in the  $P2_12_12_1$  space group). We detail the approach required to solve the novel structure of *E. coli* mutase, utilizing the CCP4 programs (Collaborative Computational Project, Number 4, 1994) *AMoRe* (Navaza, 1994), *DM* and *DM\_Multi* (Cowtan, 1994) to place electron density into the new cell and determine the phases.



**Figure 1**  
The reaction catalysed by UDP-galactopyranose mutase is unprecedented in biochemistry. The interconversion between the two ring forms involves no net redox reaction, although a flavin is involved.

Our laboratory has studied this protein because it is vital to the formation of the cell wall and or the capsule in many pathogenic bacteria. The biosynthesis of the cell wall is an attractive target for structure-assisted drug design of novel antibacterial drugs mainly because there are a large number of compounds in the cell wall that are not found in mammalian and other higher organisms. One such compound is galactofuranose (Galf). Galf is a component of the LPS in many Gram-negative bacteria, found in the O antigens of many species including *Klebsiella pneumonia* (Koplin *et al.*, 1997) and *E. coli* (Nassau *et al.*, 1996). Galactofuranosyl residues are also an important component of the arabinogalactan complex that forms part of the mycolitic layer in the cell walls of mycobacteria (Weston *et al.*, 1998). Galf is incorporated into cell walls from UDP-Galf, formed by the enzyme UDP-galactopyranose mutase (mutase) from UDP-galactopyranose (Galp) (Fig. 1; Weston *et al.*, 1998). The chemistry involved in this ring-contraction mechanism is completely unprecedented and is the source of great interest.

## 2. Results

### 2.1. Protein purification

The purification of *E. coli* mutase has been described previously (McMahon *et al.*, 1999). Production of the SeMet

derivative protein was carried out by transfection of the Met<sup>-</sup> strain B834 (Novagen) with the plasmid containing the *E. coli* gene (Nassau *et al.*, 1996) and cell growth and induction of protein production were carried out in essentially the same manner as the native. Purification of the SeMet protein was as described for the native protein, with 2 mM fresh DTT in all buffers and solutions.

### 2.2. Crystallization

**2.2.1. P<sub>21</sub> crystal form.** Crystals in the P<sub>21</sub> form were grown in 20% PEG 4K, 12% 2-propanol, 0.01 M L-cysteine, 100 mM HEPES pH 7.6 and 8 mg ml<sup>-1</sup> protein as described previously (McMahon *et al.*, 1999). SeMet protein crystals were grown in an identical manner to the wild-type protein, yielding smaller crystals than the wild type. A key point was that several seeding steps were required to obtain crystals of sufficient size for diffraction. This protocol was extremely reproducible.

**2.2.2. Re-obtaining the P<sub>21</sub>2<sub>1</sub>2<sub>1</sub> crystal form.** A solitary single crystal was obtained in the space group P<sub>21</sub>2<sub>1</sub>2<sub>1</sub> form and this was originally described in McMahon *et al.* (1999). We have been unable to reproduce this crystal. The initial crystal was grown from an early purification run of mutase and had impurities in it. Subsequent purifications resulted in homogeneous protein. Deliberately 'contaminated' protein did not crystallize. However, by the streak-seeding of P<sub>21</sub> crystals into drops containing 10–20% PEG 6K, 100 mM HEPES pH 7.0, 0.01 M L-cysteine, 2 mM DTT and 2–3 mg ml<sup>-1</sup> protein, either pre-equilibrated overnight (for the 10% drops) or seeded immediately (for the 20% drops), we were successful in growing crystals in the P<sub>21</sub>2<sub>1</sub>2<sub>1</sub> form. These crystals grew overnight to 0.2 × 0.2 × 0.1 mm in size. Repeated seedings and macroseeding with single crystals generally failed to increase the size.

**Table 1**  
Summary of crystallographic data.

Values in parentheses refer to the highest resolution shell.

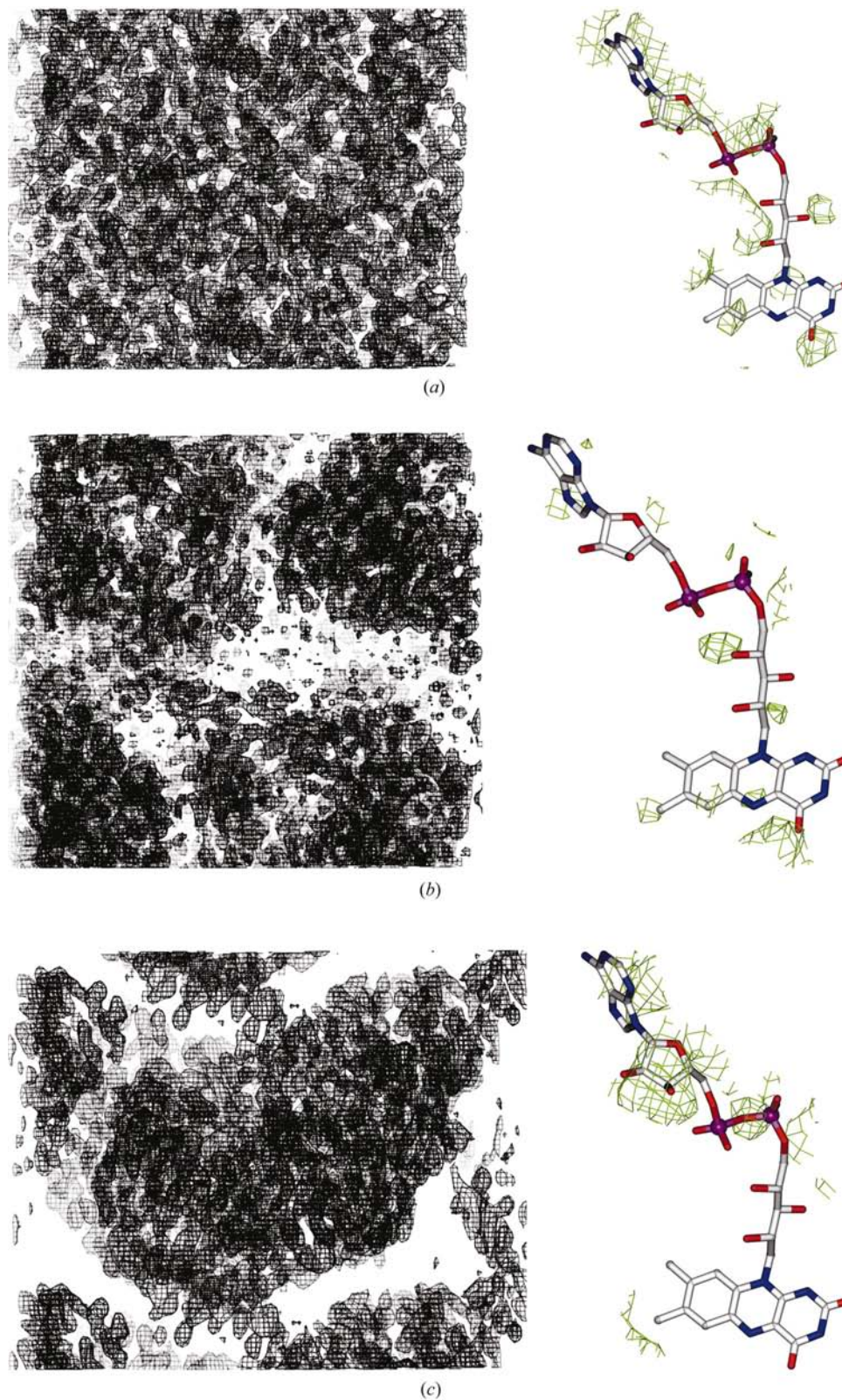
	Native data set (P <sub>21</sub> )	SeMet-1 data set (P <sub>21</sub> )†	Native data set (P <sub>21</sub> 2 <sub>1</sub> 2 <sub>1</sub> )	MAD data set (P <sub>21</sub> )		
Wavelength (Å)	0.984	0.926	0.934	λ <sub>1</sub> = 0.9794	λ <sub>2</sub> = 0.9796	λ <sub>3</sub> = 0.9393
Resolution (Å)	40–2.75	40–2.8	40–2.4	30–2.8	30–2.8	30–3.2
Highest shell (Å)	2.85–2.75	2.9–2.8	2.52–2.4	2.9–2.8	2.9–2.8	3.3–3.2
Unit-cell parameters (Å)	a = 68.8, b = 57.4, c = 97.3, γ = 97	a = 65.1, b = 53.9, c = 95.5, α = β = 90, γ = 98	a = 56.6, b = 98.1, c = 132.5, α = β = γ = 90	a = 69.5, b = 57.0, c = 97.3, α = β = 90, γ = 96.4		
Unique reflections	19034	15510	28288	35522	35938	24113
I/σ(I)	5.8 (0.9)	17.8 (3.9)	10.9 (3.3)	10.6 (2.5)	9.1 (1.3)	7.6 (1.7)
Average redundancy	2.9 (2.8)	2.4 (2.5)	3.9 (3.7)	1.7 (1.7)	1.7 (1.7)	1.7 (1.7)
Completeness‡ (%)	95 (51)	93 (93)	98 (93)	97 (96)	98 (96)	99 (98)
Anomalous completeness‡ (%)	—	93 (93)	—	97 (96)	98 (96)	97 (96)
R <sub>merge</sub> ‡§	9.0 (39.5)	5.5 (32.3)	5.9 (22.4)	8.1 (31.7)	7.8 (55.4)	15.0 (56.8)
f <sup>+</sup> /f <sup>-</sup>	—	—	—	-6.2/6.0	-10.2/3.06	-3.02/4.4
Mosaic spread	0.73	1.9	0.5	1.7	1.7	1.9
No. of sites	—	—	—	9	—	—
R <sub>iso</sub> ¶ relative to native (P <sub>21</sub> )	N/A	0.525	N/A	0.13	0.14	0.17
Figure of merit	—	—	—	0.43	—	—

† SeMet-1 data set collected as described in McMahon *et al.* (1999) on only a single wavelength. ‡ Anomalous, completeness, redundancy and R<sub>merge</sub> calculations for MAD data sets treat F<sup>+</sup> and F<sup>-</sup> as separate observations. § R<sub>merge</sub> = ∑ I(h)<sub>j</sub> = I(h)/∑ ∑ I(h)<sub>j</sub>, where I(h) is the measured diffraction intensity and the summation includes all observations. ¶ R is the R factor = ∑ |F<sub>o</sub> - ∑ |F<sub>c</sub>|/∑ |F<sub>o</sub>|. R<sub>free</sub> is the R factor calculated using 5% of the data that were excluded from the refinement.

### 2.3. Data collection

**2.3.1.  $P2_1$  data.** The SeMet-1 data set to 3.0 Å was collected as described previously (McMahon *et al.*, 1999). The native and SeMet-2 data sets for the  $P2_1$  crystal form were collected on beamline ID14-4 at the ESRF synchrotron in Grenoble. Although crystals were generally highly variable in diffraction and had high mosaic spread, several crystals of sufficient quality were found to collect the necessary data sets. Owing to the fragile nature of the crystals, we used a poor-quality larger crystal to EXAFS scan in order to determine the three wavelengths for the MAD data set. The native data set consisted of 180 non-overlapping 1° images collected for 30 s to 2.7 Å and the SeMet data set was to 2.8 Å resolution for two wavelengths and 3.2 Å for the third (Table 1), each consisting of 180 1° non-overlapping images collected for 15 s. Data were processed using the programs *MOSFLM* (Leslie, 1992) and *SCALA* (Evans, 1997), treating the Bijvoet pairs as independent measurements for the SeMet data set.

**2.3.2.  $P2_12_12_1$  data.** The data for the  $P2_12_12_1$  crystal form were collected on beamline ID14-1 at the ESRF synchrotron in Grenoble. A single crystal of about 0.15 × 0.15 × 0.075 mm in size had to be cryoprotected in a relatively complex manner. The crystal was moved (using a glass capillary ~0.2 mm size) into a stabilizing buffer (25% PEG 4K, 15% 2-propanol, 100 mM HEPES pH 7.6) and after ~1 min was moved rapidly through stabilizing buffer plus 7.5% glycerol into stabilizing buffer plus 15% glycerol, in which it was then frozen. The initial image of this crystal showed high mosaicity. The mosaicity was significantly reduced by a round of reannealing/flash-freezing by blocking the Cryostream for 5 s. A data set was collected to 2.4 Å resolution,



**Figure 2**

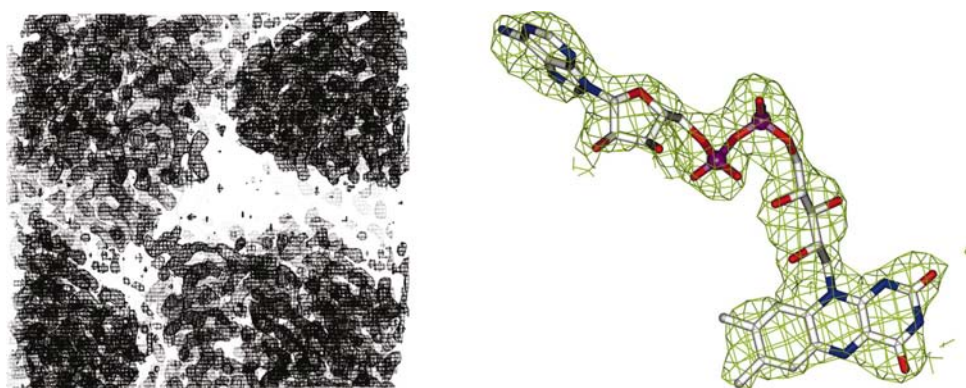
Experimental electron density contoured at  $1\sigma$ . On the left is density of the same 75 Å<sup>2</sup> region of space, while on the right is a stereoview of FAD with the density from the map. Figures generated with Snapshot from *O* (Jones *et al.*, 1991) (left) and *BOBSCRIPT* (Esnouf, 1997) utilizing the *GL\_RENDER* interface (L. Esser and J. Deisenhofer, unpublished work) and *POV-Ray* (right). (a) Initial map from *SOLVE*. (b) Map after initial solvent flattening, averaging and phase extension. (c) Map after mask improvement, cross-crystal averaging with native, SIROAS and non-isomorphous SeMet-1 data. Back-translation of the final refined structure into the  $P2_1$  cell shows that the solvent boundaries in (b) are completely wrong.

consisting of 220  $0.5^\circ$  non-overlapping images collected for 30 s each. Data were processed using *MOSFLM* and *SCALA* (details are given in Table 1).

## 2.4. Structure solution

**2.4.1.  $P2_1$  solution.** The program *SOLVE* 1.17 (Terwilliger & Berendzen, 1999) was used to locate the selenium sites in the  $P2_1$  crystal form. Nine of a possible 14 sites were found ( $Z$  score = 20, FOM = 0.43). Eight of these sites were consistent with a twofold rotation suggested by the self-rotation map. The initial map was improved by solvent flattening with *DM* (Cowtan, 1994) with phase extension to 2.7 Å. As shown in Fig. 2(b), the map was very poor and no convincing evidence of secondary structure was found. Close analysis of the data indicated that the anomalous signal from the SeMet data was very weak. One indicator of the poor quality of the anomalous signal is the ratio  $\Delta_{\text{ano}}/\sigma(\Delta_{\text{ano}})$ , which in the case of the mutase data was significantly less than one for all three wavelengths (0.67, 0.62 and 0.58 for the peak, inflection and remote wavelengths, respectively). *SOLVE* also indicated the same problem. Some additional 'improvement' of the phases was made by placing a 50 Å sphere as a mask centred at the centroid of the selenium positions. This mask was improved by manually rebuilding the map to fill areas of density through visual inspection.

*MLPHARE* (Otwinowski, 1991) was used to generate SIROAS phases using the peak wavelength for SeMet-2 against the native data set. Consistent with the weak signal, the anomalous occupancy tended to zero when refined in *MLPHARE*. We then began cross-crystal averaging between the native (SIROAS phases), SeMet-2 (MAD phases) and SeMet-1 data. A monomer mask derived from the earlier 50 Å mask was used as the NCS mask. The translation matrices for these data sets were assumed to be unity and allowed to refine in the program. This is despite effectively complete non-isomorphism between SeMet-1 and the two other data sets; the average mean fractional isomorphous difference was 0.525. Attempts to improve the matrices prior to *DMMULTI*



**Figure 3**

Final experimental density map for the  $P2_12_12_1$  data. Density is contoured at  $1\sigma$  and is of the same region of space as in Fig. 2. This figure is of a different cell ( $P2_12_12_1$ ) compared with Fig. 2 ( $P2_1$ ) and so cannot be directly compared.

**Table 2**

*AMoRe* results for molecular replacement using density.

Solution	<i>a</i>	<i>b</i>	<i>c</i>	<i>x</i>	<i>y</i>	<i>z</i>	Corr. coeff.	<i>R</i> factor
Solution 1	41.09	86.47	337.52	0.1623	0.1780	0.0908	24.7	52.2
Solution 2	91.90	82.46	182.15	0.0902	0.2498	0.000	20	54.6
Solution 3	137.23	90.00	156.92	0.3437	0.1806	0.4177	19.3	53.5
Solution 4	26.96	43.32	35.55	0.9988	0.2981	0.0330	8.3	57.5
Solution 1 after fitting	41.02	86.67	337.85	0.1630	0.1756	0.0908	26.1	51.9

by density-placement calculations (Cowtan, 1994) gave worse results (as measured by correlation coefficients and visual examination of maps). The results from this *DMMULTI* run were promising, with a final correlation coefficient for monomer *A* between native and SeMet-1 of 0.935 and an FOM of 0.66 for 2.8 Å  $P2_1$  native data. This map was the used for an initial model-building step. Clear secondary-structure elements (mostly  $\alpha$ -helices with a few  $\beta$ -strands) were visible for the first time in the map. The map is shown in Fig. 2(c). A polyalanine model consisting of 300 residues was traced into this map, possible secondary-structure elements were assigned and this model was used to search *DALI* (Holm & Sanders, 1993) and *DEJAVU* (Kleywegt & Jones, 1997) for similar proteins; however, no solutions were found. An attempt was made to refine this model using *CNS* (Brunger *et al.*, 1998), but the model did not refine.

**2.4.2.  $P2_12_12_1$  solution.** At the point where the  $P2_1$  data failed to render a solution, the  $P2_12_12_1$  crystal form was rediscovered and the data was collected. The transformation matrix between the two crystal forms was generated by using molecular placement of the cross-crystal averaged electron density from  $P2_1$ . The  $P2_1$  model was used to create a mask of the monomer inside the  $P2_1$  unit cell (using *MAPMASK*; Collaborative Computational Project, Number 4, 1994). This mask was then applied to the 'final' map from the same solution  $P2_1$  solution (NCS, cross-crystal averaged) (using *MAPMASK* and *OVERLAPMASK*; Jones & Stuart, 1991). The region outside of the mask was flattened and the resulting

density was transformed to a  $P1$  symmetry cell (*EXTEND*; Collaborative Computational Project, Number 4, 1994). This map was then back-transformed using *SFALL* (Collaborative Computational Project, Number 4, 1994) to generate the structure factors, which were normalized to *E* values (*ECALC*; Collaborative Computational Project, Number 4, 1994) and ordered for use in *AMoRe* (Navaza, 1994) by *CAD* (Collaborative Computational Project, Number 4, 1994). The calculated data were sorted and tabled in *AMoRe* and then run against the  $P2_12_12_1$  data to find

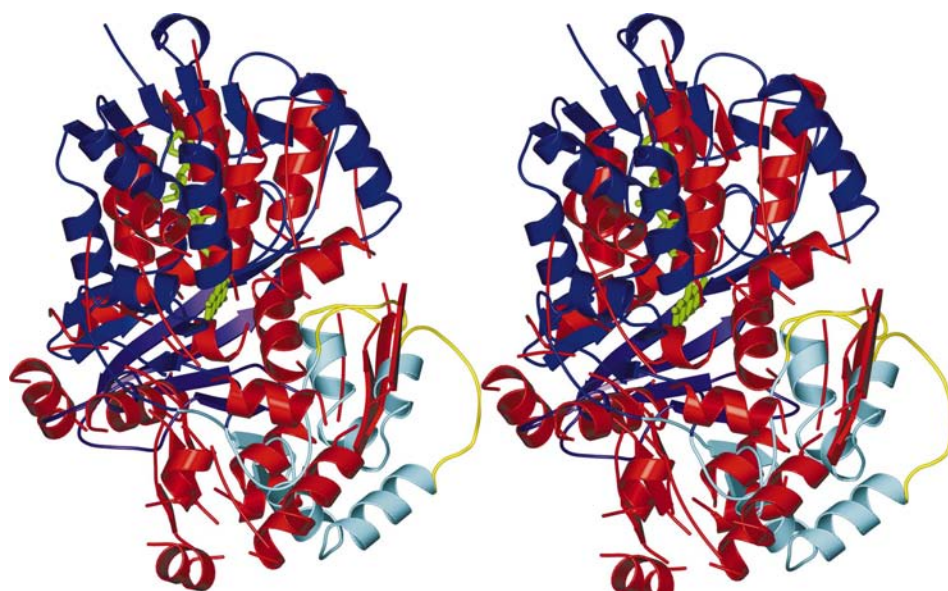
**Table 3**

Results of *DMMULTI* cross-crystal averaging between  $P2_1$  and  $P2_12_12_1$  crystal forms.

Data set	Correlation coefficient		FOM	
	Before <i>DM</i> †	After <i>DM</i> †	Before	After
$P2_1$ native	−/0.19	−/0.92	0.13	0.62
SeMet-1	0.00/0.03	0.88/0.86	0.00	0.61
$P2_12_12_1$	0.01/0.01	0.84/0.80	0.00	0.48

† Each crystal has two monomers, monomer *A* and monomer *B*. The starting point for averaging is monomer *A* of  $P2_1$  native. The values refer to the correlation between  $P2_1$  monomer *A* and monomer *A*/monomer *B* of the other crystals.

solutions for the monomer (see Table 2). The matrix between the two monomers in the  $P2_12_12_1$  unit cell was derived by applying the *AMoRe* monomer solution to the positions of the Se atoms determined from *SOLVE*. The relationship between the two sets of Se atoms determined the matrix between the monomers. We based this on the assumption that the position of the two monomers did not change between the two different crystal forms. We then used cross-crystal averaging (*DMMULTI*; Cowtan, 1994) using this transformation and four data sets [ $P2_1$  native with the 'final' phases,  $P2_1$  SeMet-2 peak wavelength with phases from *SOLVE/DM* (SIROAS phases), SeMet-1 data set and the  $P2_12_12_1$  data set]. The rotation matrix for the different  $P2_1$  data sets was assumed to be unity and all of the matrices were allowed to be refined by *DMMULTI*. Table 3 shows the starting and finishing correlation coefficients and FOMs from the final *DMMULTI* run. Fig. 3 shows the density of the experimental phases (prior to refining) at the position of refined FAD.

**Figure 4**

Superposition between model built from  $P2_1$  data (in red, unrefinable) and final refined mutase monomer from  $P2_12_12_1$  data (in various blues). Superposition performed initially with *LSQKAB* (Kleywegt & Jones, 1997) and then adjusted manually in *O* (Jones *et al.*, 1991).

**Table 4**

Final refinement statistics for *E. coli* mutase.

No. of protein atoms	6084
No. of water molecules	321
No. of heteroatoms	106
Root-mean-square deviations from	
Ideal bond lengths†	0.012
Ideal bond angles†	1.4
Mean temperature factor	
Main chain	26.55
Side chain	29.11
Heteroatoms	19.37
Water molecules	32.23
Crystallographic <i>R</i> factor (%)	18.52
Free crystallographic <i>R</i> factor‡ (%)	24.52

† R.m.s. deviation from Engh and Huber ideal values (Engh & Huber, 1991). ‡  $R_{\text{free}}$  is calculated on 5% of data excluded during refinement.

At this point, a polyaniline model was built into the experimental density map, allowing tracing of 321 residues (of 367); the FAD was also found. The resulting model and known NCS operators were then used for real-space averaging (*RAVE*). This allowed further identification of 20 residues and also allowed the joining of different segments of the structure. This model was run through *DALI* (Holm & Sanders, 1993) and *DEJAVU* (Kleywegt & Jones, 1997). This confirmed that the FAD-binding domain was similar to a number of other proteins; however, the second domain and linking region were novel. Refinement was begun using *CNS* (Brunger *et al.*, 1998), starting with simulated annealing, rigid-body refinement and positional/*B*-factor refinement. This allowed the location of the final 26 residues and joining of the last segments (Fig. 4). Further refinement proceeded smoothly and resulted in a model with final refinement statistics as listed in Table 4.

### 3. Discussion

The solution of the *E. coli* mutase structure has proven to be exceedingly difficult owing to the small size and high degree of mosaicity of the crystals. The  $P2_1$  crystal form with native and SeMet derivative (MAD) data sets proved insufficient to solve the phase problem, most likely owing to the very poor anomalous signal in the MAD data sets. The poor anomalous signal was probably the result of the high mosaicity and concomitant poor diffraction. Attempts to obtain heavy-metal derivatives of the  $P2_1$  crystal form were hampered by our inability to screen crystals in-house. Limited synchrotron time also prevented extensive screening and no useful information was obtained

with the putative platinum derivative collected at Grenoble (McMahon *et al.*, 1999).

The discovery that the  $P2_12_12_1$  crystals could be grown by seeding with the  $P2_1$  crystal form proved instrumental in solving the structure. The initial  $P2_12_12_1$  crystal reported earlier was probably grown in a drop containing enough impurities to provide a nucleation site; however, subsequent attempts to return to this condition were unsuccessful owing to, ironically, greater attention to sample purity. Micro/streak seeding of  $P2_1$  crystals into these drops was enough to allow the  $P2_12_12_1$  form to be grown. These crystals proved to be much better for diffraction purposes, despite a much more rigorous freezing protocol.

Owing to the fact that there is apparently a mathematical relationship between the two cells (as in Table 1:  $a_{P2_12_12_1} = b_{P2_1}$ ,  $b_{P2_12_12_1} = c_{P2_1}$ ,  $c_{P2_12_12_1} = 2a_{P2_1}$ ), we began deriving the rotation matrix between the two crystal forms mathematically. The rapid success of molecular replacement using density meant that we never completed the mathematical transformation.

During a post-mortem analysis we tried to place electron density from the initial  $P2_1$  MAD data directly obtained from *SOLVE*. No solution was found, although careful examination of the rotation-function solutions did show one peak with approximately the correct rotation/translation solution, although the  $z$ -axis translation was quite different (solution = 42.07, 87.17, 332.57, 0.21, 0.18, 0.38). The same test of the density-modified map (Fig. 2*b*) gave similarly poor results. Experience in the laboratories suggests a good indicator of a correct solution can be found by viewing the min/max density of the rotation map. The ratio of (max. density/r.m.s. deviation):(min. density/r.m.s. deviation) gives a fairly reliable indicator of height of peaks above noise level. In our example, the successful molecular replacement gave a ratio of 8.3:(−3.9) compared with 4.9:(−0.9) and 5.0:(−3.9) for the results from *SOLVE* and *DM* (Cowtan, 1994), respectively.

Our results confirm the adage that while correlation coefficient and FOM are useful guides toward determining whether the phases derived are correct, visual examination of maps and the appearance of such easily discernable features as  $\alpha$ -helices are better guides towards structure solution. We have also demonstrated that placement of electron density and cross-crystal averaging are very powerful tools for solving structures using very poor initial phases. The post-mortem analysis shows that only by extracting the maximum phase information from the SeMet-2 data (by manual adjustment of

the mask and SIROAS phases from the peak wavelength) and cross-crystal averaging against the non-isomorphous SeMet-1 data set (which was very poor quality), could good enough density be obtained for placement in the  $P2_12_12_1$  cell.

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## References

- Brunger, A. T., Adams, P. D., Clore, G. M., DeLano, W. L., Gros, P., Grosse-Kunstleve, R. W., Jiang, J.-S., Kuszewski, J., Nilges, M., Pannu, N. S., Read, R. J., Rice, L. M., Simonson, T. & Warren, G. L. (1998). *Acta Cryst.* **D54**, 905–921.
- Collaborative Computational Project, Number 4 (1994). *Acta Cryst.* **D50**, 760–763.
- Cowtan, K. (1994). *Jnt CCP4/ESF-EACBM Newsl. Protein Crystallogr.* **31**, 34–38.
- Engh, R. A. & Huber, R. (1991). *Acta Cryst.* **A47**, 392–400.
- Esnouf, R. M. (1997). *J. Mol. Graph. Model.* **15**, 132–134.
- Evans, P. R. (1997). *Jnt CCP4/ESF-EACBM Newsl. Protein Crystallogr.* **33**, 22–24.
- Holm, L. & Sanders, C. (1993). *J. Mol. Biol.* **233**, 123–138.
- Jones, T. A., Zou, J.-Y., Cowan, S. W. & Kjeldgaard, M. (1991). *Acta Cryst.* **A47**, 110–119.
- Jones, Y. & Stuart, D. I. (1991). *Proceedings of the CCP4 Study Weekend. Isomorphous Replacement and Anomalous Scattering*, edited by W. Wolf, P. R. Evans & A. G. W. Leslie, pp. 39–48. Warrington: Daresbury Laboratory.
- Kleywegt, G. J. & Jones, T. A. (1997). *Methods Enzymol.* **277**, 525–545.
- Koplin, R., Brisson, J.-R. & Whitfield, C. (1997). *J. Biol. Chem.* **272**, 4121–4128.
- Leslie, A. G. W. (1992). *Jnt CCP4/ESF-EACBM Newsl. Protein Crystallogr.* **26**.
- McMahon, S. A., Leonard, G. L., Buchanan, L. V., Giraud, M.-F. & Naismith, J. H. (1999). *Acta Cryst.* **D55**, 399–402.
- Nassau, P. M., Martin, S. L., Brown, R. E., Weston, A., Monsey, D., McNeil, M. R. & Duncan, K. (1996). *J. Bacteriol.* **178**, 1047–1052.
- Navaza, J. (1994). *Acta Cryst.* **A50**, 157–163.
- Otwinowski, Z. (1991). *Proceedings of the CCP4 Study Weekend. Isomorphous Replacement and Anomalous Scattering*, edited by W. Wolf, P. R. Evans & A. G. W. Leslie, pp. 80–86. Warrington: Daresbury Laboratory.
- Terwilliger, T. C. & Berendzen, J. (1999). *Acta Cryst.* **D55**, 849–861.
- Weston, A., Stern, R. J., Lee, R. E., Nassau, P. M., Monsey, D., Martin, S. L., Scherman, M. S., Besra, G. S., Duncan, K. & McNeil, M. R. (1998). *Tubercle Lung Dis.* **78**, 123–131.